

M1 EEA-RM-GBM

EMEAM2FM - BIOCHIMIE ET BIOMATÉRIAUX

I – Les protéines

- 1- Les acides aminés
 - A- Structure générale
 - B- Propriétés acido-basiques et optiques
- 2- Les peptides
 - A- Définitions
 - B- Structure primaire
 - C- La liaison peptidique
 - D- Exemples de peptides
- 3- La structure des protéines

II – Les Enzymes

- 1 - Introduction – Définitions
- 2 - Les cofacteurs enzymatiques
 - A - biotine (ou vitamine B8)
 - B - Nicotinamide Adénine Dinucléotide (NAD^+)
- 3 - La réaction enzymatique
 - A - réaction non-catalysée
 - B - catalyse enzymatique
 - C - notion de site actif
 - D - introduction à la cinétique enzymatique
 - E - mesures enzymatiques : quantification d'une biomolécule

IV – Les Acides Nucléiques

- 1- Bases azotées
 - A- 2 sortes de bases : purines et pyrimidines
 - B- Absorbance dans l'U.V.
 - C- Densité de charge
- 2- Nucléosides et nucléotides
 - A- Liaison avec 2 types de sucres
 - B- Modification avec l'acide phosphorique
 - C- Nomenclature
- 3- Structures spatiales
 - A- Association des nucléotides dans un acide nucléique
 - B- Complémentarité des bases
 - C- Double hélice/Propriétés
 - D- Modifications chimiques des acides nucléiques
- 4- Des nucléotides remarquables : ATP, AMPc et GMP
- 5- Séquençage de l'ADN et PCR

V – Les lipides

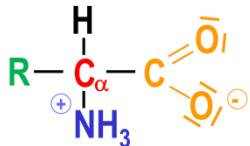
- 1- Introduction
- 2- Nature et propriétés des acides gras
- 3- Lipides contenant des acides gras
- 4- Lipides dérivés d'acides gras
- 5- Lipides issus d'unités isopréniques
- 6- Purification des lipides

VI – Les glucides

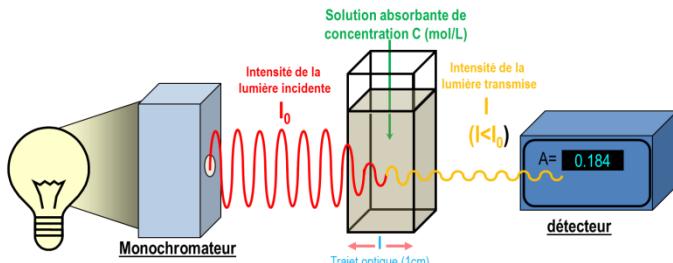
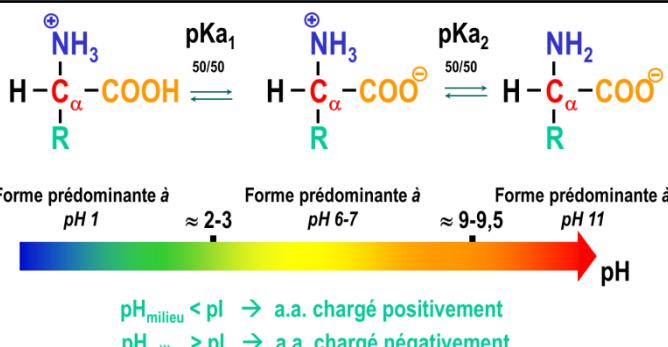
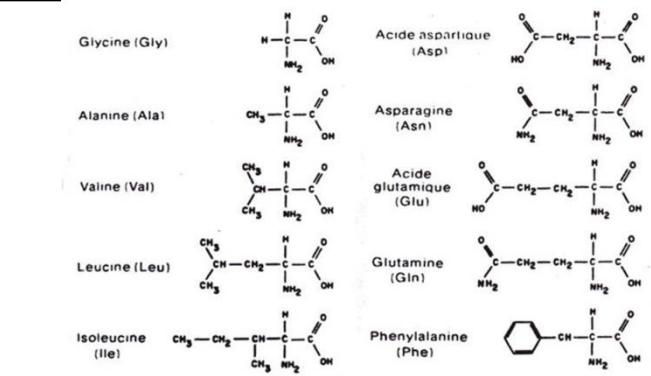
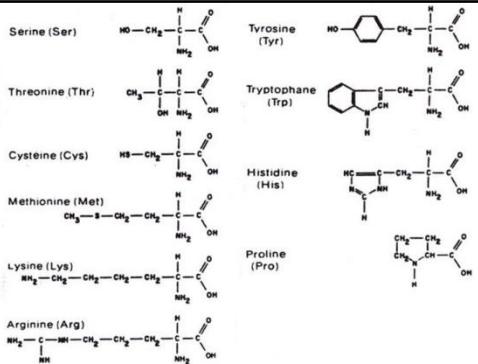
- 1- Introduction
- 2- Monosaccharides et disaccharides
- 3- Polysaccharides
- 4- Glycoprotéines et glycolipides
- 5- Dérivés d'oses
- 6- Détection du glucose : glycémie

III – Techniques de Purification et d'Analyse

- 1 - Solubilisation – extraction des protéines
- 2 - Précipitation différentielle
 - A - précipitation isoélectrique
 - B - précipitation par des sels
- 3 - Techniques chromatographiques
 - A - échange d'ions
 - B - exclusion / diffusion
 - C - affinité
- 4 - Techniques électrophorétiques
 - A - électrophorèse sur papier
 - B - électrophorèse sur gel de polyacrylamide
- 5 - Technique immunoenzymatiques

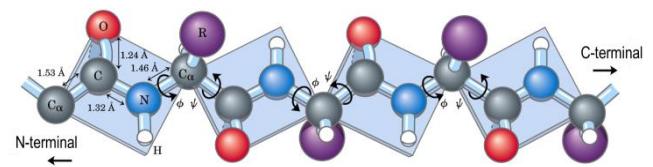
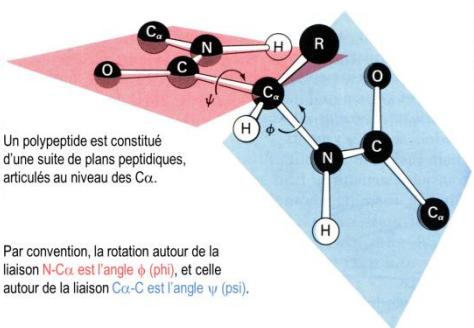
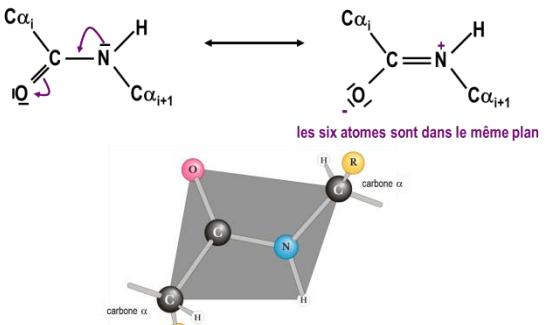
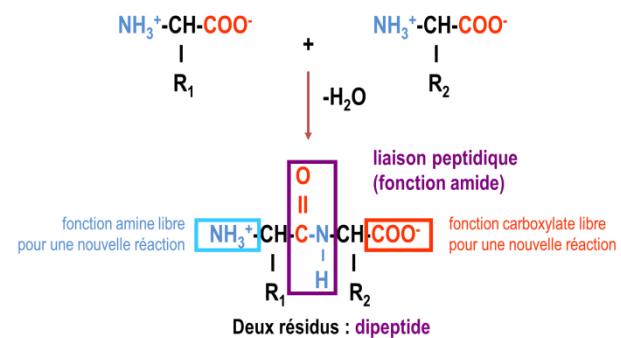
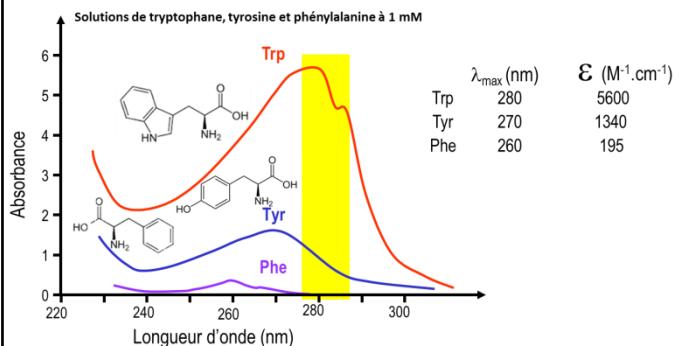


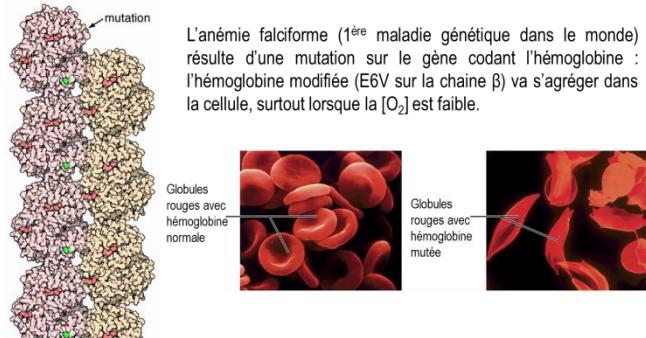
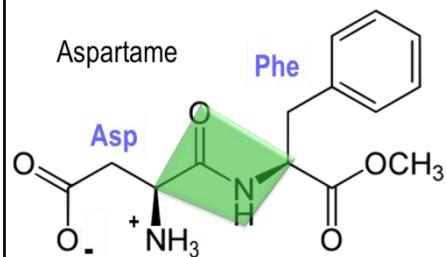
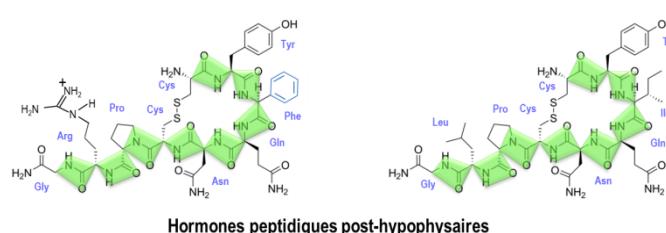
Symbole	Code 3 lettres	Nom
A	Ala	Alanine
C	Cys	Cystéine
D	Asp	Aspartate
E	Glu	Glutamate
F	Phe	Phénylalanine
G	Gly	Glycine
H	His	Histidine
I	Ile	Isoleucine
K	Lys	Lysine
L	Leu	Leucine
M	Met	Méthionine
N	Asn	Asparagine
P	Pro	Proline
Q	Gln	Glutamine
R	Arg	Arginine
S	Ser	Sérine
T	Thr	Thréonine
V	Val	Valine
W	Trp	Tryptophane
Y	Tyr	Tyrosine



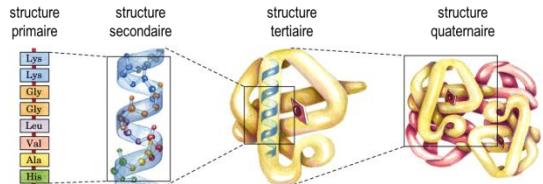
- La fraction de la lumière absorbée par une solution à une longueur d'onde donnée dépend :

- 1- de l'épaisseur de la solution que la lumière doit traverser (trajet optique)
- 2- de la concentration de la solution en espèces absorbantes

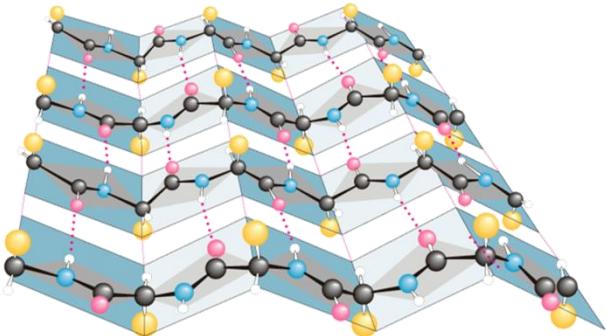
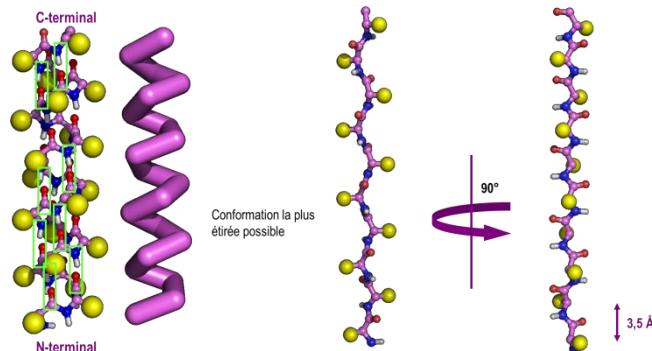




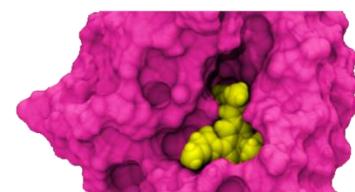
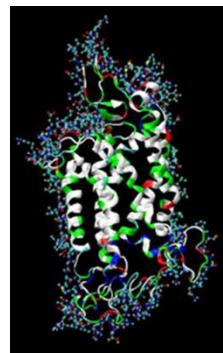
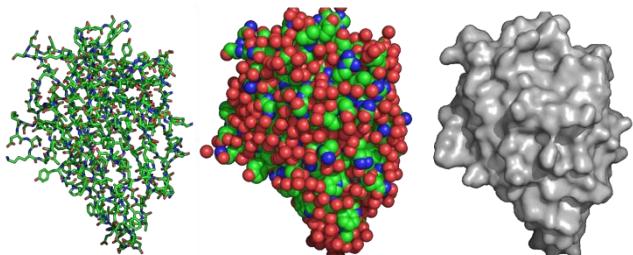
Différents niveaux de structuration pour une protéine



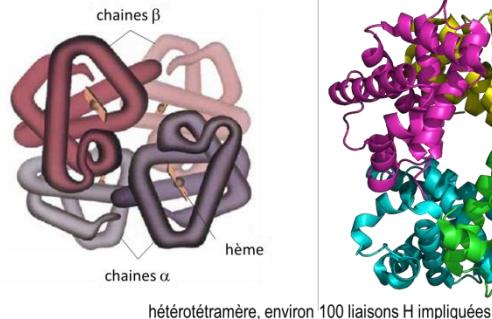
Structure primaire : enchaînement des acides aminés (séquence)
 Structure secondaire : repliement local de la chaîne polypeptidique
 Structure tertiaire : repliement global de la chaîne polypeptidique
 Structure quaternaire : assemblage de plusieurs chaînes polypeptidiques



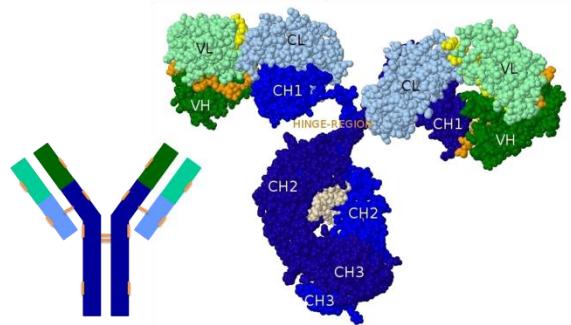
La concanavaline A

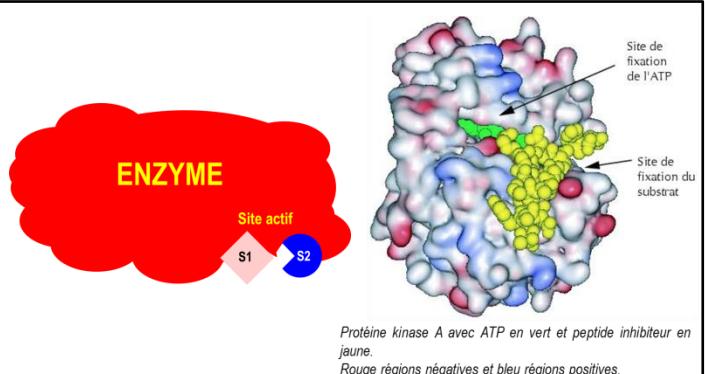
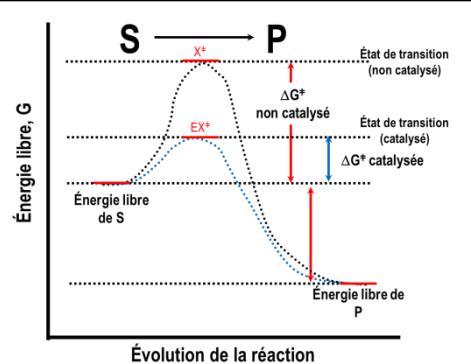
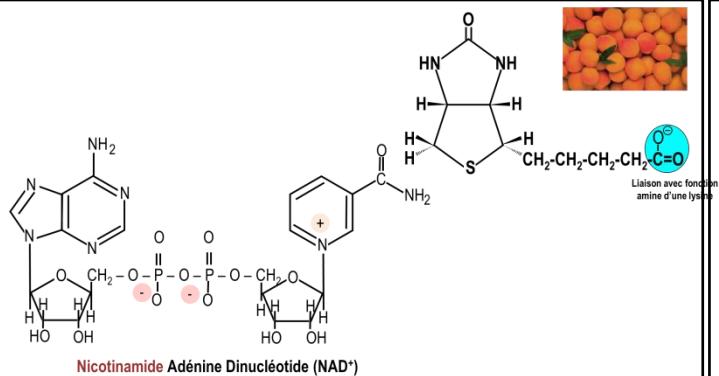


L'hémoglobine



Les immunoglobulines

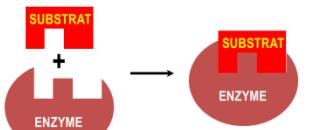




1894 MODELE DE FISCHER : CLE-SERRURE



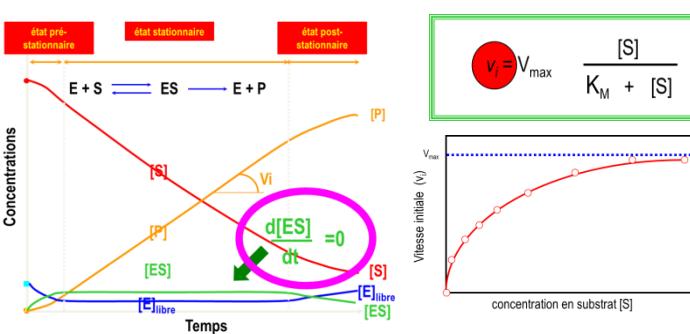
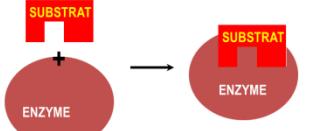
Hermann Emil Fischer



1958 MODELE DE KOSHLAND : AJUSTEMENT INDUIT

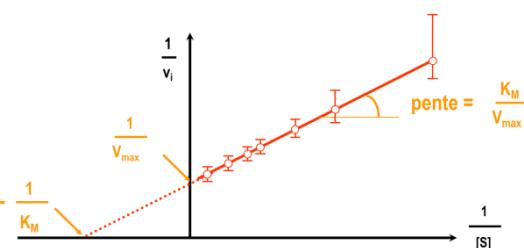


Daniel Koshland Jr.



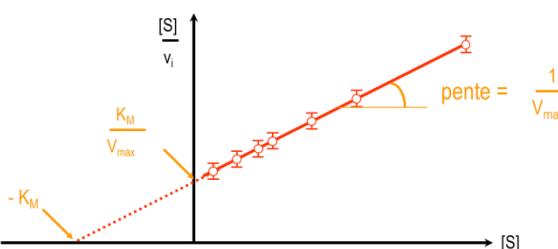
Lineweaver - Burk

$$\frac{1}{V_i} = \frac{K_M}{V_{max}} \times \frac{1}{[S]} + \frac{1}{V_{max}}$$



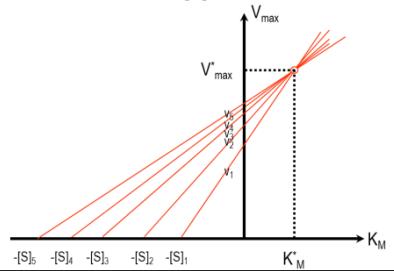
Hanes

$$\frac{[S]}{V_i} = \frac{K_M}{V_{max}} + \frac{1}{V_{max}} \times [S]$$

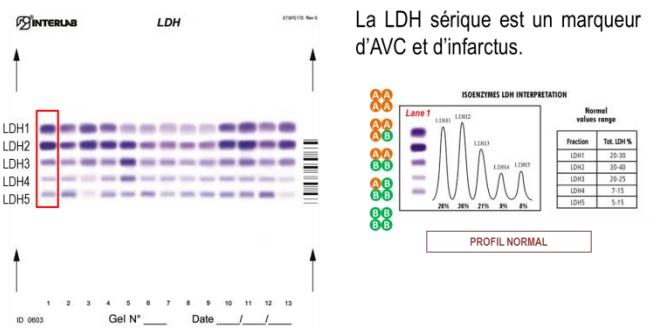
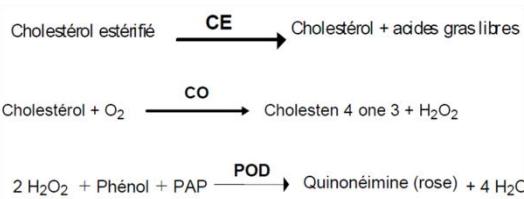


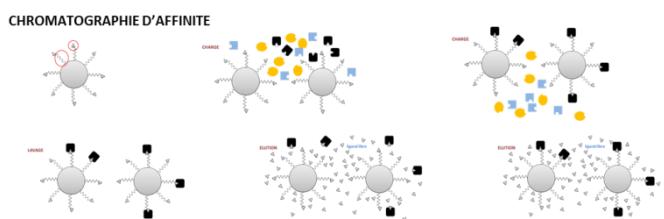
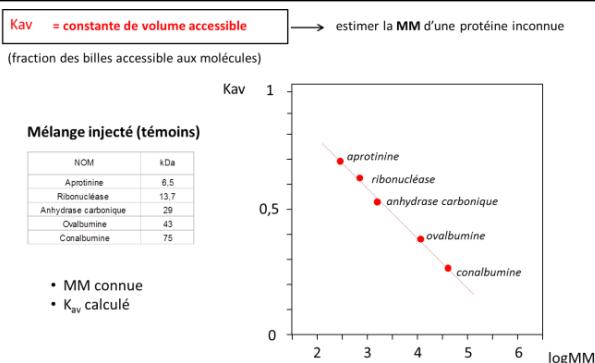
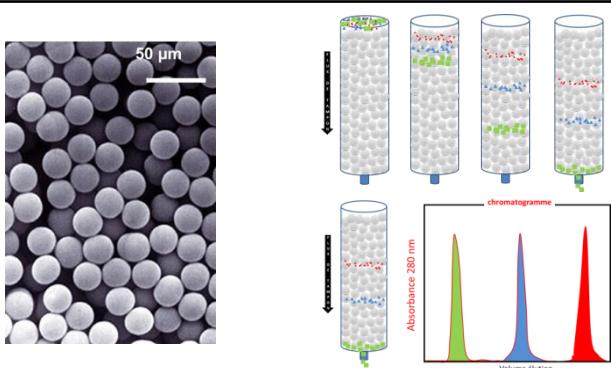
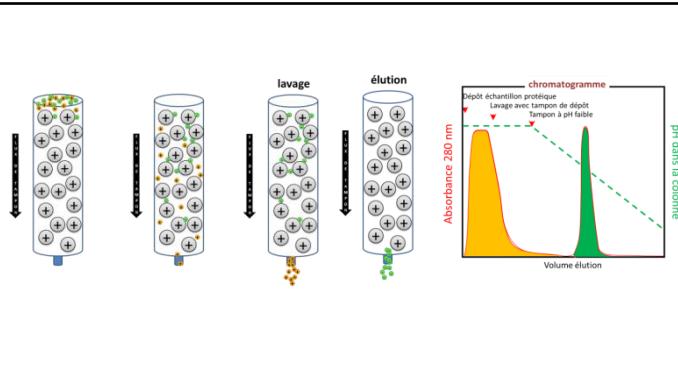
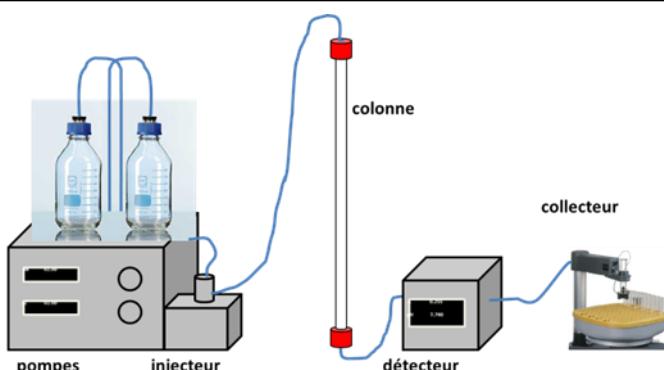
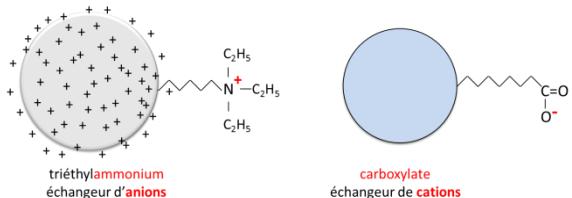
Eisenthal - Cornish-Bowden

$$\frac{V_{max}}{[S]} = V_i + \frac{V_i}{V_{max}} \times \frac{K_M}{[S]}$$

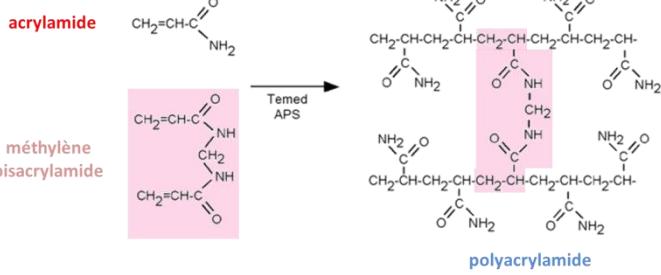
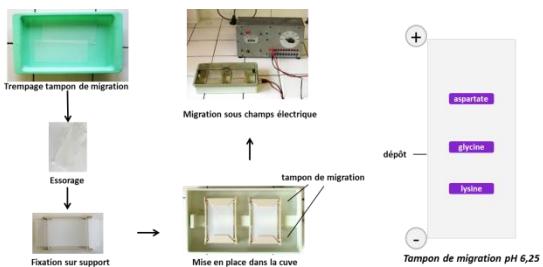


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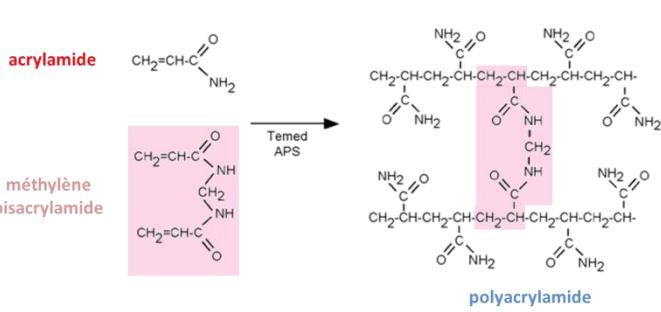
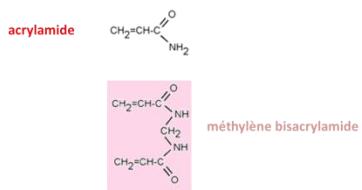




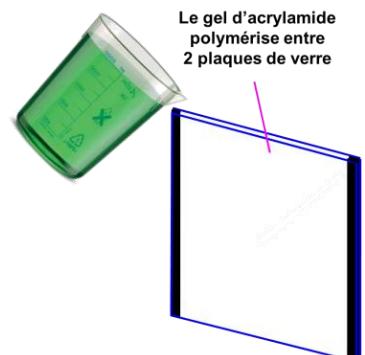
ELECTROPHORESE SUR PAPIER



ELECTROPHORESE SUR GEL ACRYLAMIDE



Acrylamide/bisacrylamide
Tampon Tris pH6,8 ou 8,8
SDS
Persulfate d'ammonium
TEMED

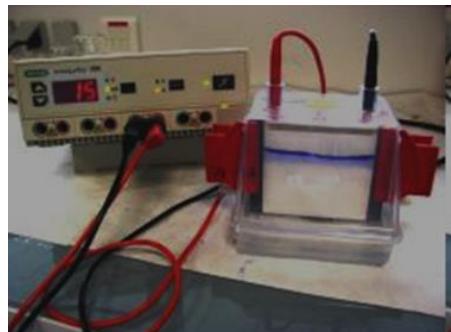
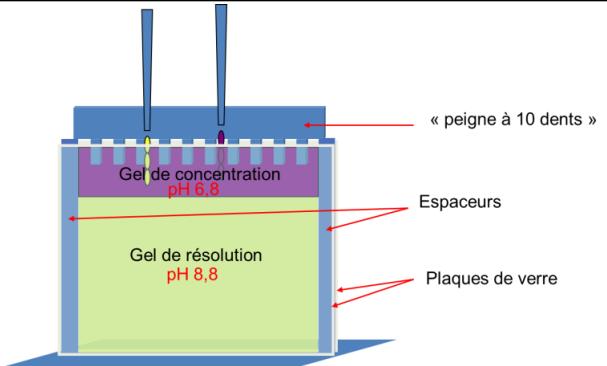
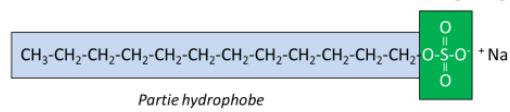


Préparation des échantillons protéiques

Les protéines sont incubées en présence de :

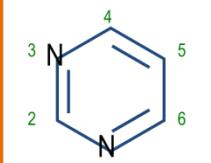
- détergent anionique (SDS : sodium dodécyl sulfate)
- agent réducteur (β mercaptoéthanol) : rompt les ponts disulfure
- tampon de haute densité (glycérol) / colorant bleu.
- 5 minutes à 100°C.

Partie chargée négativement

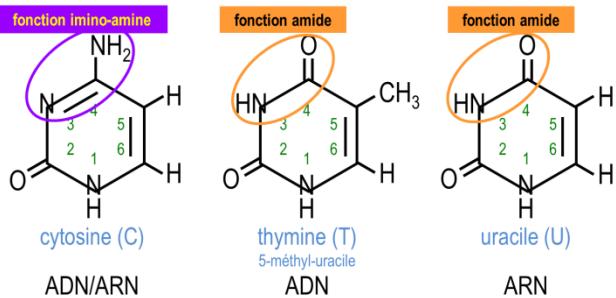
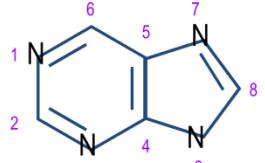


Il existe 2 sortes de bases azotées hétérocycliques :

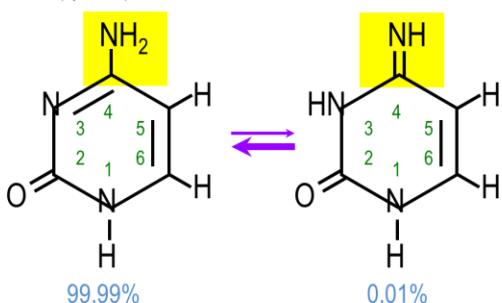
bases pyrimidiques (noyau pyrimidine)



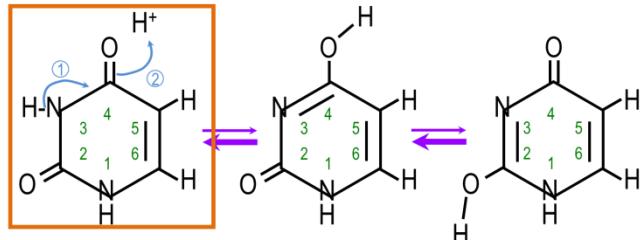
bases puriques (noyau purine)



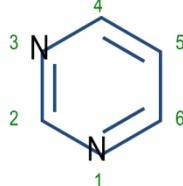
Équilibre amino-imino (cytosine)



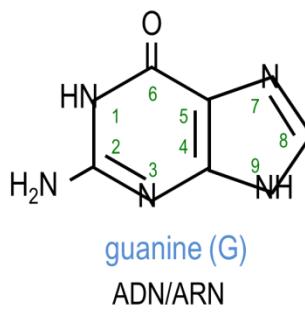
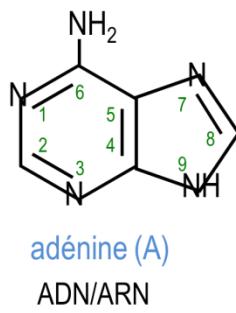
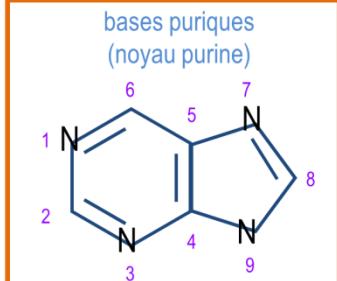
Équilibre énol-cétone (thymine-uracile)



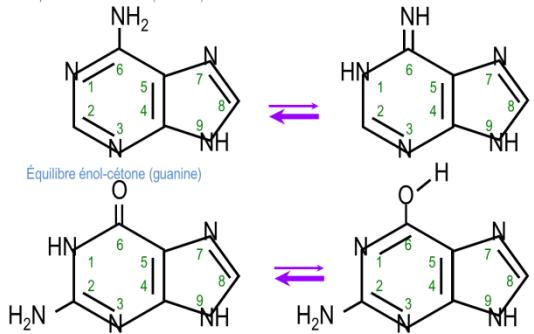
bases pyrimidiques (noyau pyrimidine)



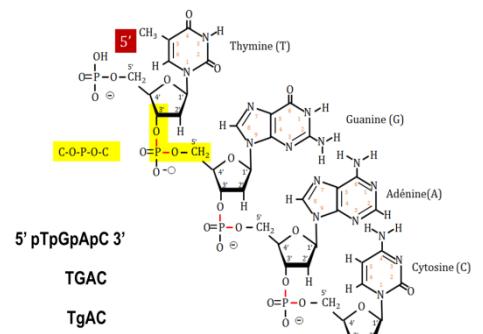
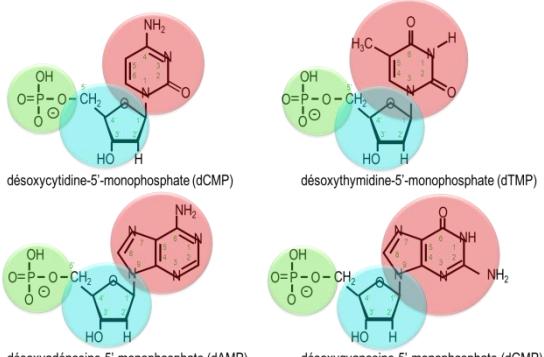
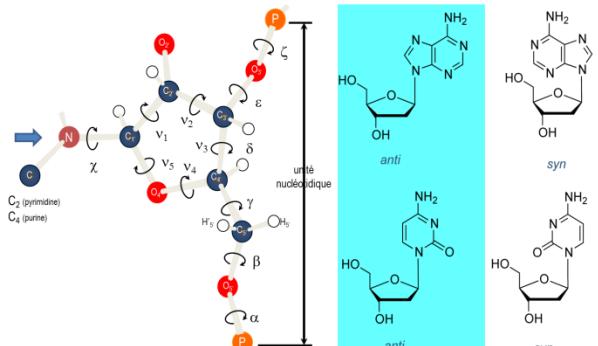
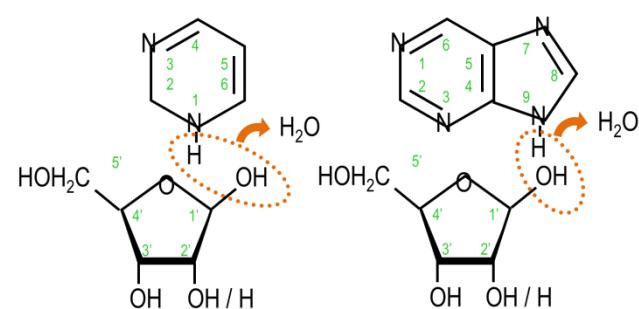
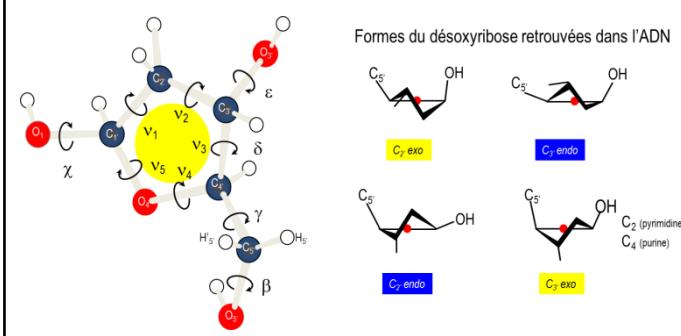
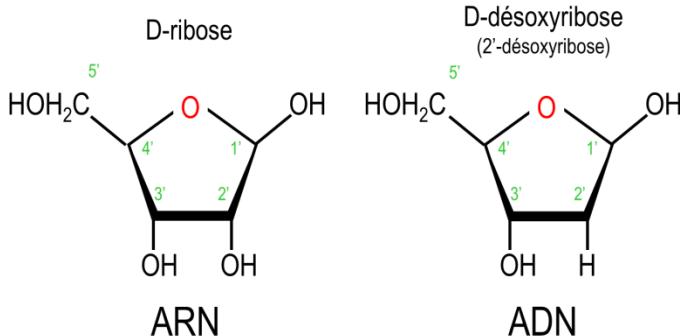
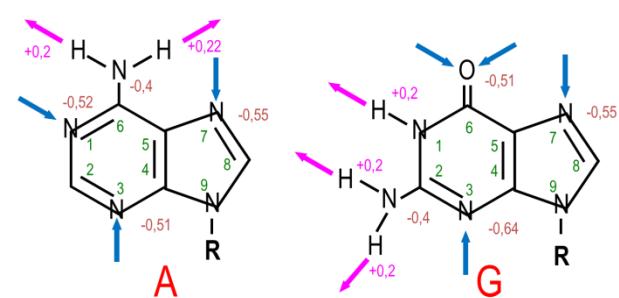
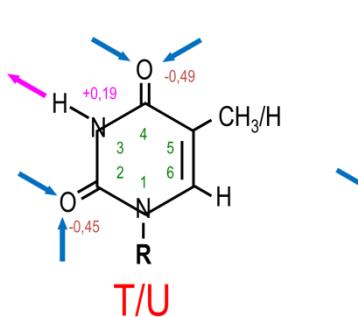
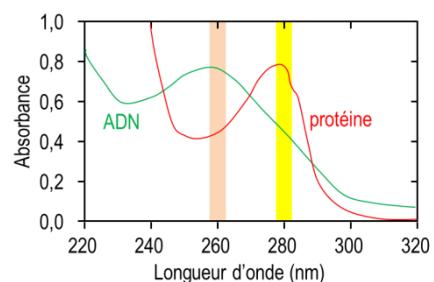
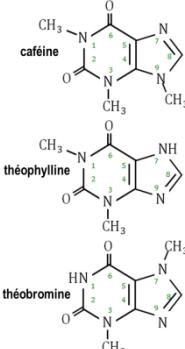
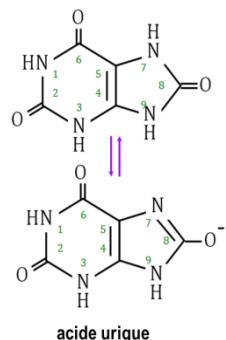
bases puriques (noyau purine)



Équilibre amino-imino (adénine)

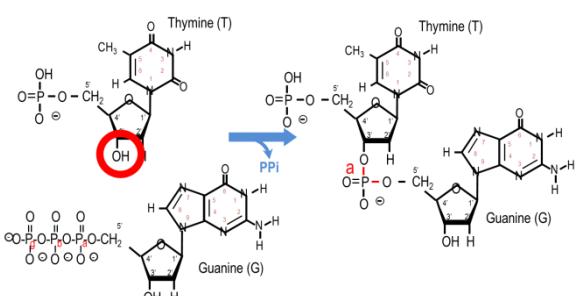
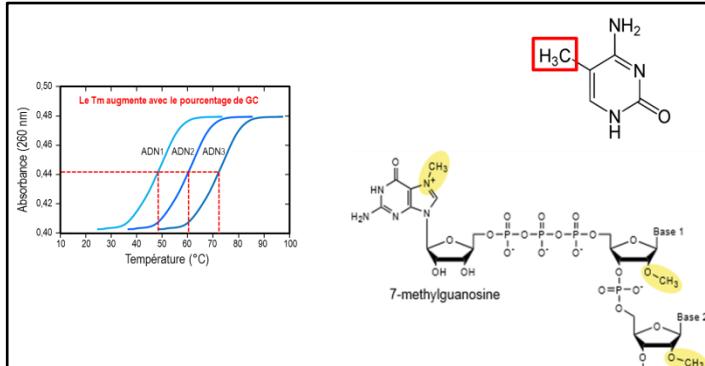
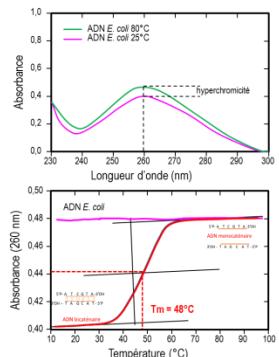
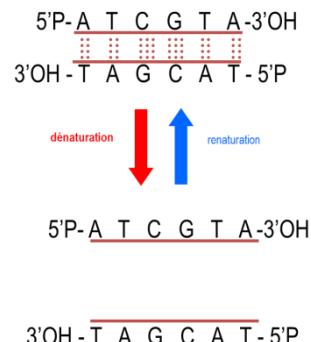
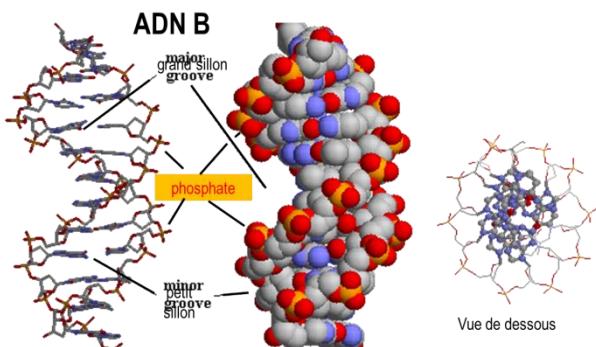
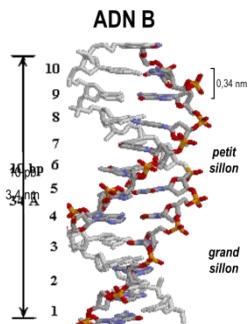
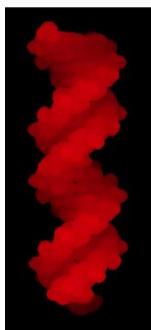
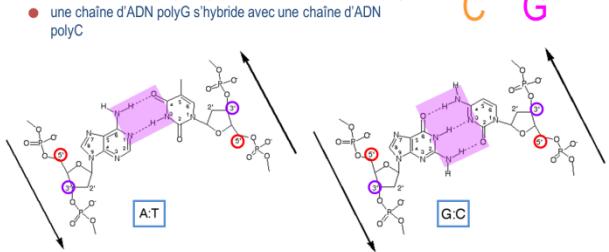


Équilibre énol-cétone (guanine)

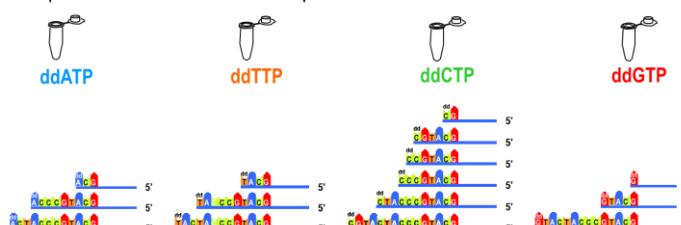


- nombre de A = nombre de T
- nombre de C = nombre de G
- une chaîne d'ADN polyA s'hybride avec une chaîne d'ADN polyT
- une chaîne d'ADN polyG s'hybride avec une chaîne d'ADN polyC

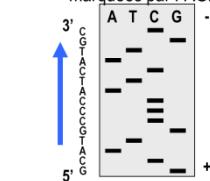
Appariement de Watson et Crick



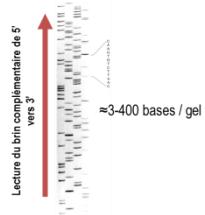
Séquences obtenues dans chaque tube.



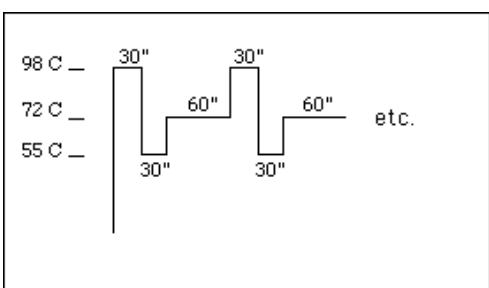
Séparation des molécules marquées par PAGE.

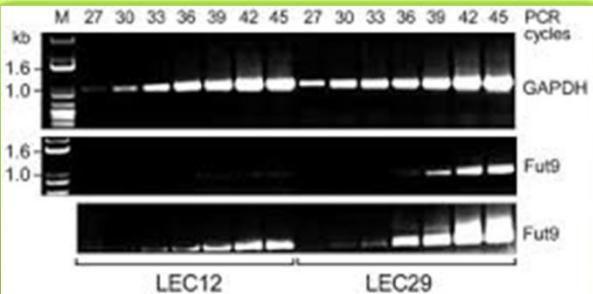


Séquence du brin complémentaire

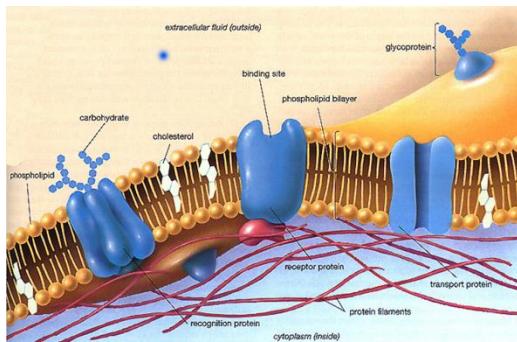
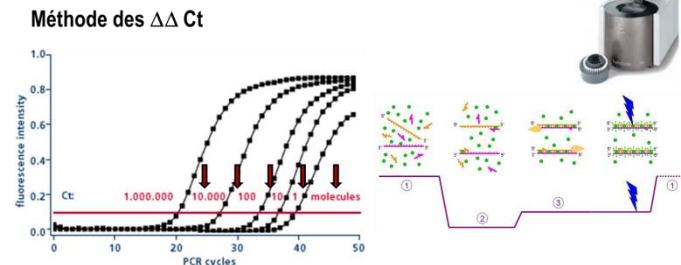


Autoradiogramme d'un gel de séquence d'ADN (polyacrylamide)

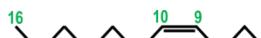




PCR en temps réel utilisant du SYBR Green comme sonde.



- Acide palmitique ($C_{16:0}$)
 - Acide n-hexadécanoïque
 - Acide stéarique ($C_{18:0}$)
 - Acide n-octadécanoïque

- Acide palmitoléique C_{16:1} Δ⁹


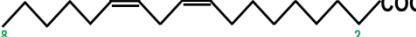
acide ω7
 - Acide oléique C_{18:1} Δ⁹


acide ω9

- Acide linoléique C_{18:2} Δ^{9,12}

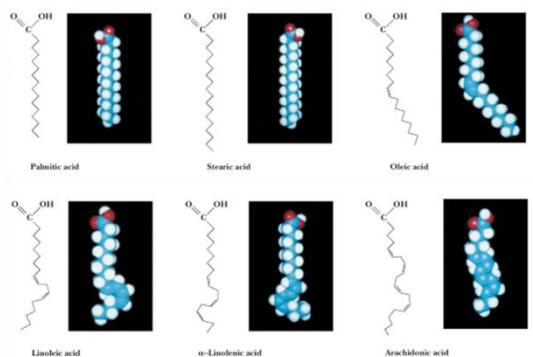
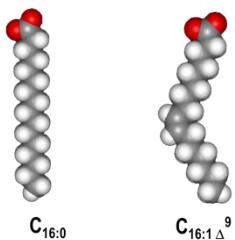


 - Acide linolénique C_{18:3} Δ^{9,12,15}

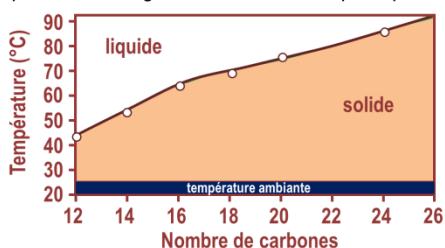

 - Acide arachidonique C_{20:4} Δ^{5,8,11,14}



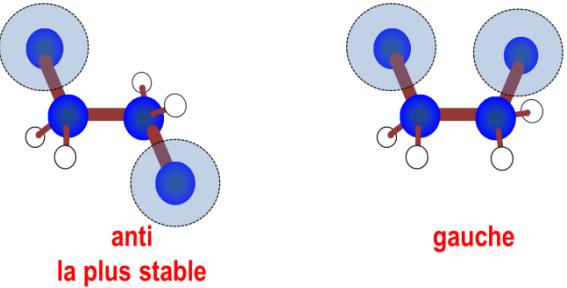
La configuration *cis* de la double liaison induit une courbure rigide de la chaîne aliphatique



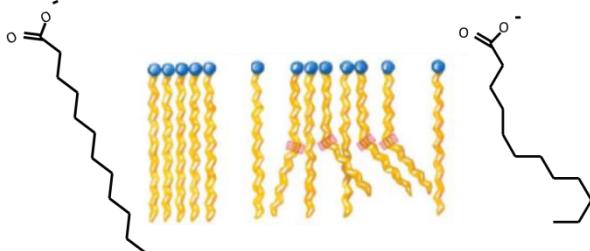
- Point de fusion : température de passage de l'état gel à l'état fluide.
 - dépend de la longueur de la chaîne aliphatique



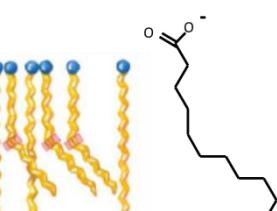
Liberté de rotation des liaisons simples C-C :
2 positions extrêmes appelées **anti** et **gauche**



Basse température : position **anti** prédominante. Les acides gras sont **ordonnés** : état **gel**



Température élevée : position **gauche** prédominante. Les acides gras sont **désordonnés** : état **fluide**

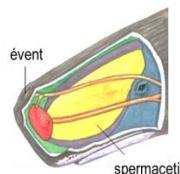


Acide gras saturé	Point de fusion (°C)	Acide gras insaturé	Point de fusion (°C)
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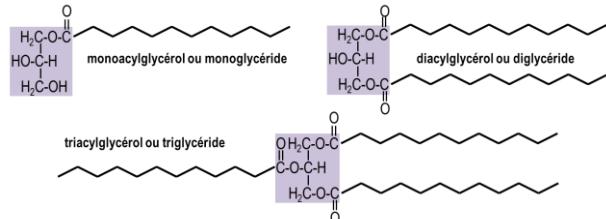
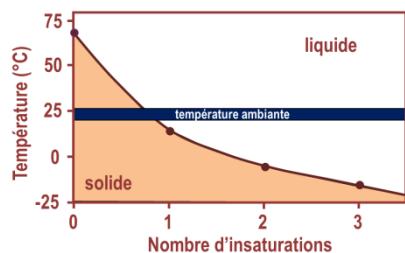
$C_{12:0}$	44,2	$C_{16:1}$	-0,5
$C_{14:0}$	53,9	$C_{18:1}$	13,4
$C_{16:0}$	63,1	$C_{18:2}$	5
$C_{18:0}$	69,6	$C_{18:3}$	-11
$C_{20:0}$	76,5	$C_{20:4}$	-49,5
$C_{22:0}$	86,0		



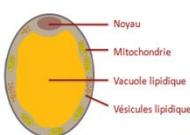
acides gras essentiels



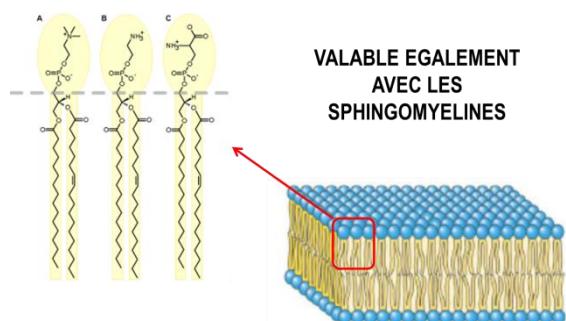
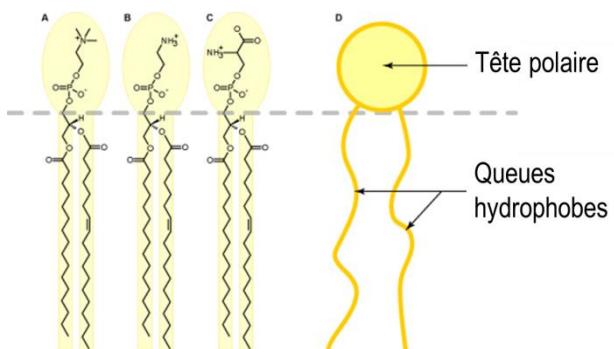
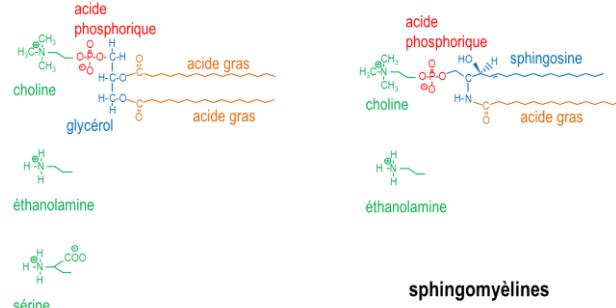
4°C → état solide et dense, l'animal plonge
37°C → état liquide et léger, l'animal remonte



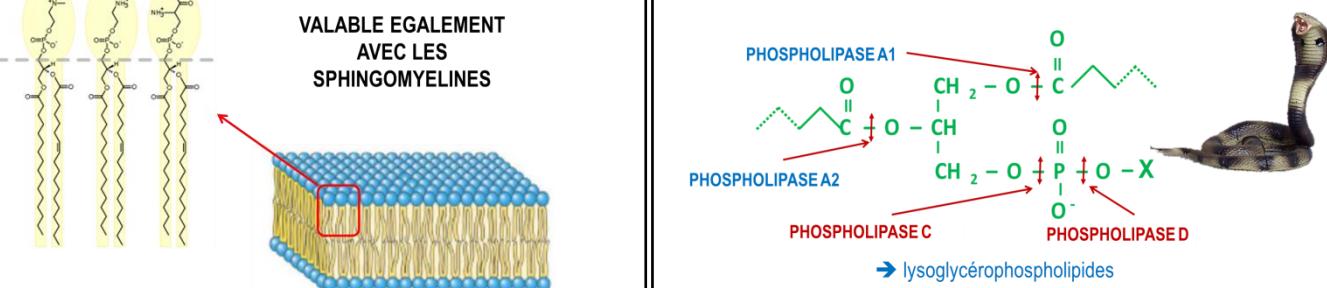
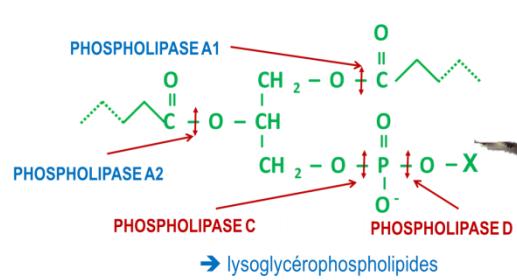
Adipocytes

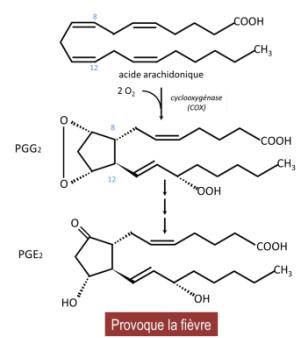
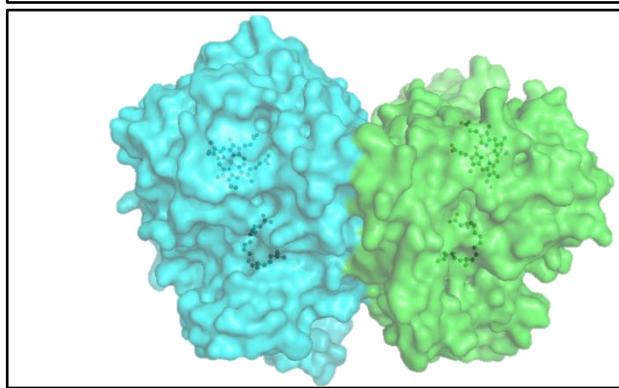
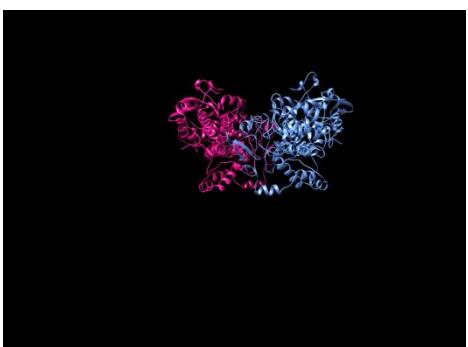
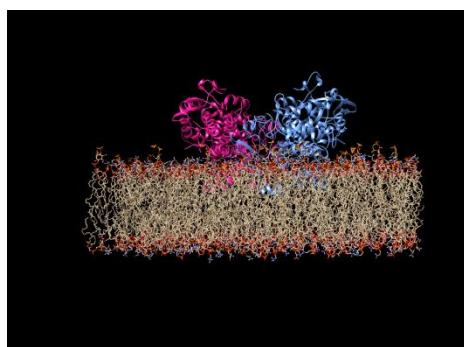
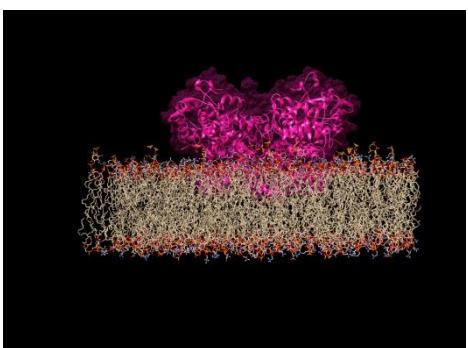
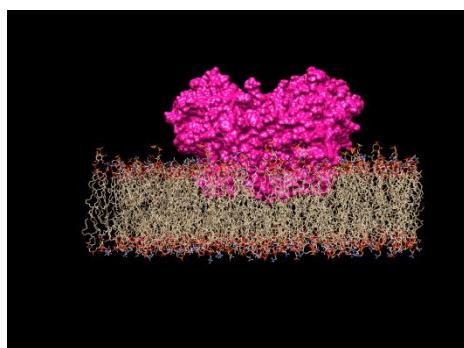
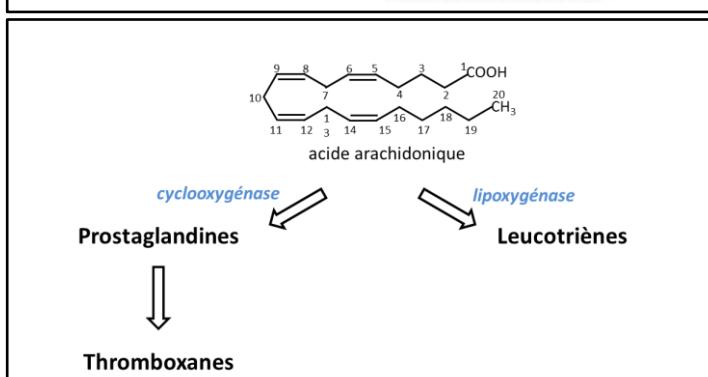
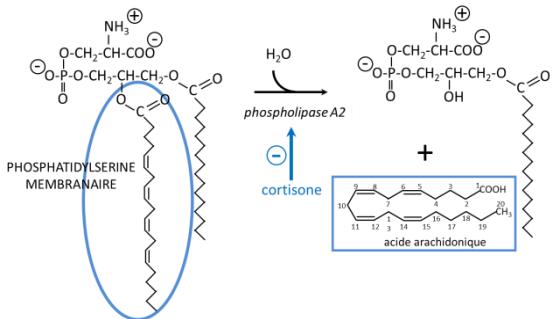
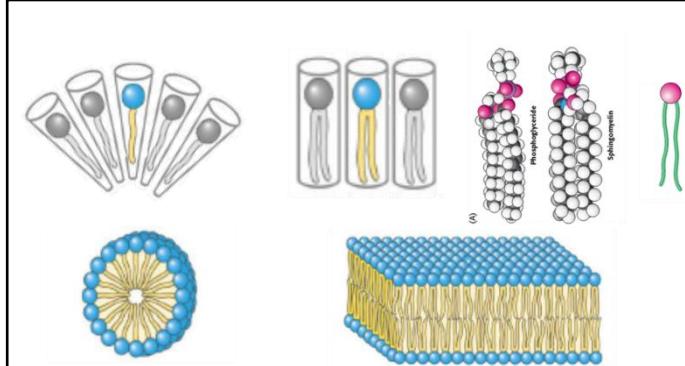


Adipocyte humain

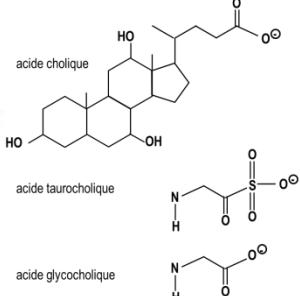
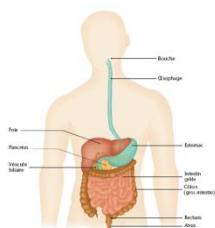
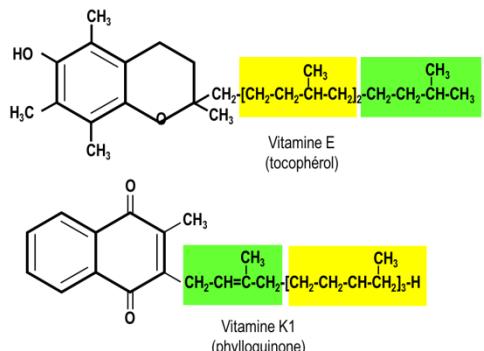
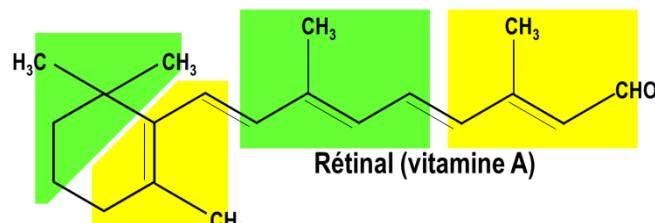
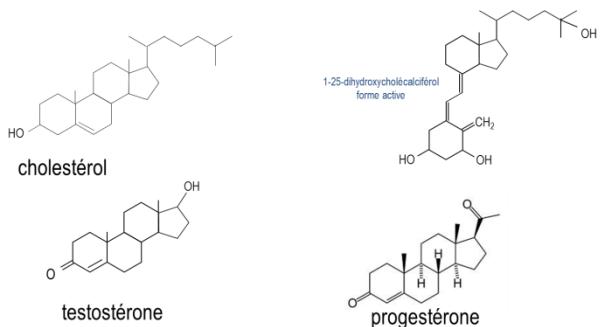
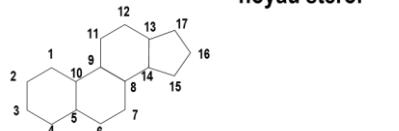


VALIDE EGALEMENT
AVEC LES
SPHINGOMYELINES





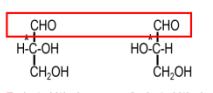
noyau stérol



1 2 3 4

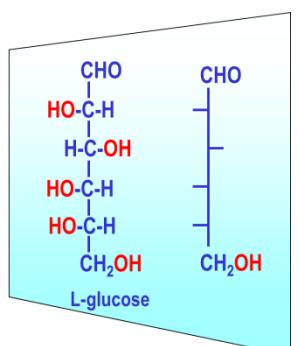
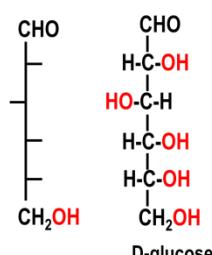
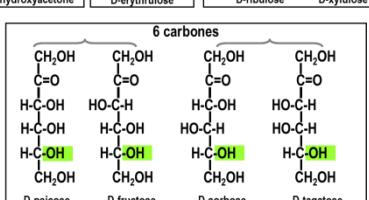
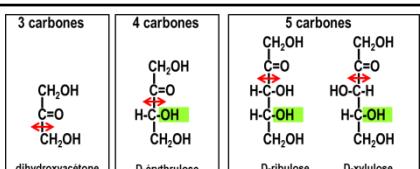
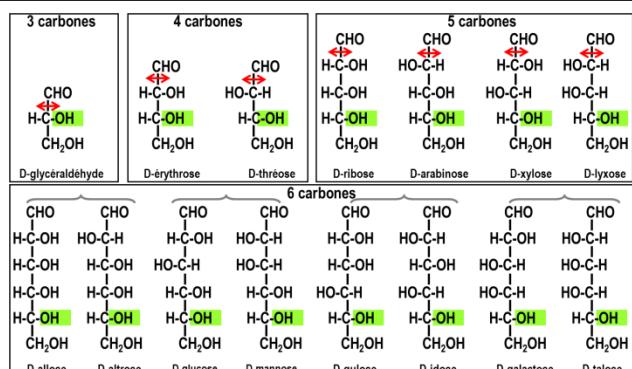
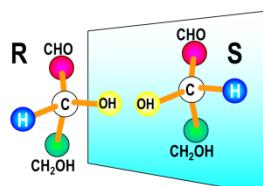
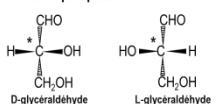
① les lipides sont déposés et se fixent sur la colonne.
Les lipides sont élus avec des solutions de plus en plus hydrophobes : ② méthanol, ③ acetonitrile, ④ acétone.

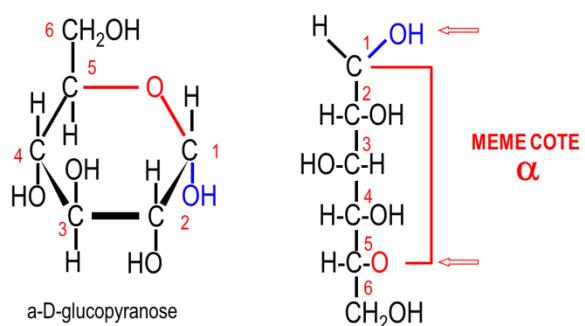
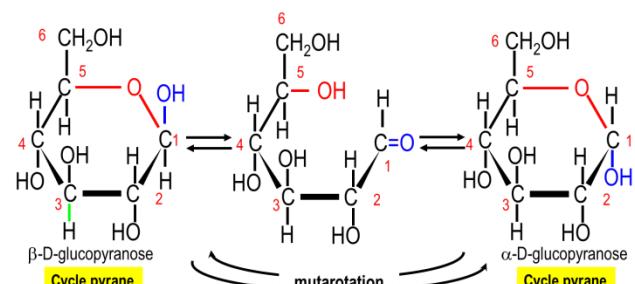
Projections de Fisher



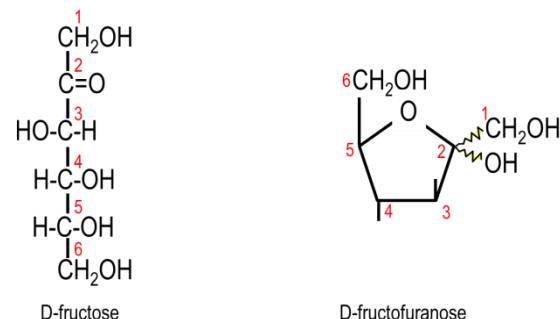
liaison horizontale ————— au dessus du plan de la feuille
liaison verticale ————— au dessous du plan de la feuille

Formules en perspective

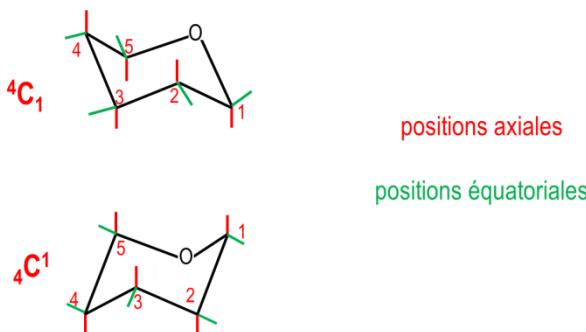
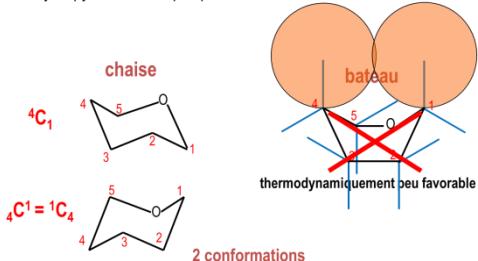




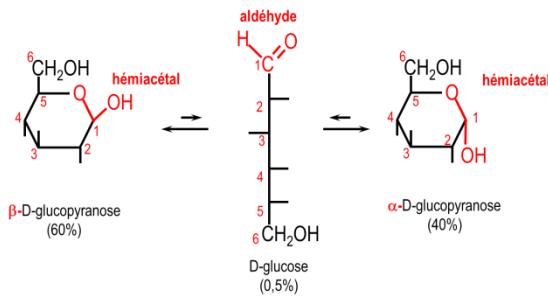
Cyclisation du fructose



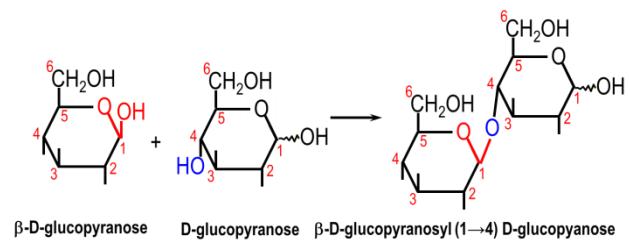
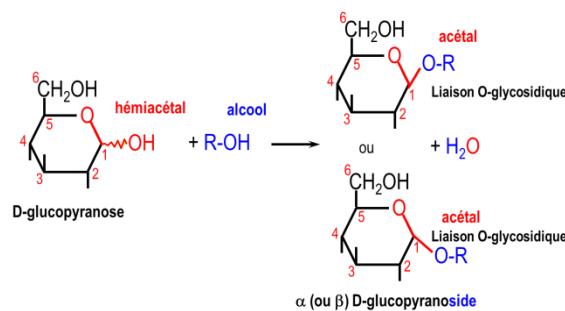
Le cycle pyranose n'est pas plan !



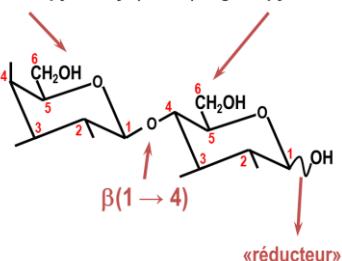
Fonction hémiacétal : mutarotation



Fonction hémiacétal : liaison osidique (ou glycosidique)

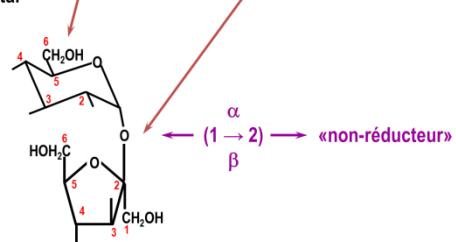


Lactose : $\beta\text{-D-galactopyranosyl (1} \rightarrow 4\text{)}\text{-D-glucopyranose}$

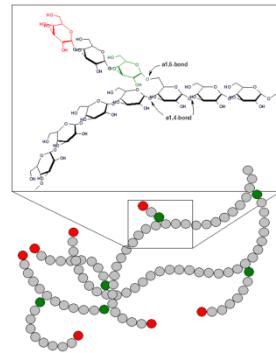
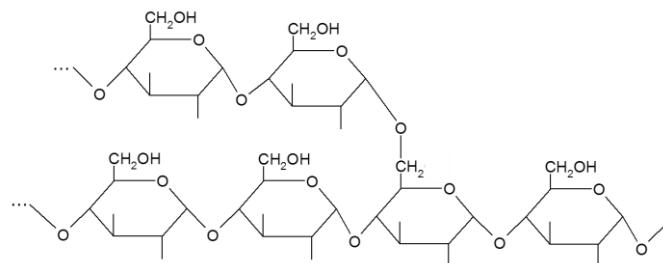


Saccharose : $\alpha\text{-D-glucopyranosyl (1} \rightarrow 2\text{)}\text{-}\beta\text{-D-fructofuranoside}$

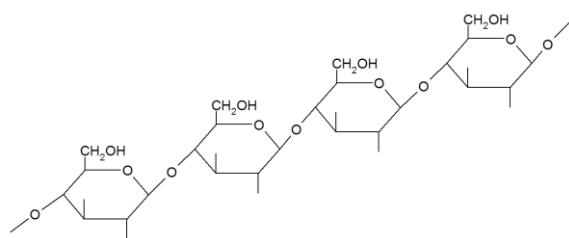
Sucre végétal



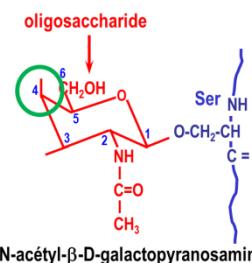
D-glucopyranoses liés entre eux par des liaisons α (1 → 4) avec des ramifications par des liaisons α (1 → 6).



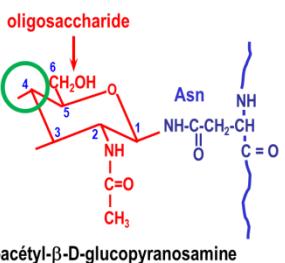
Cellulose : $(\beta\text{-D-glucopyranosyl (1 → 4)-}\beta\text{-D-glucopyranose})_n$



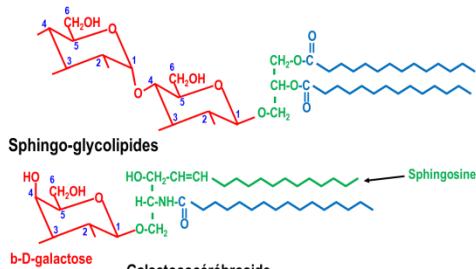
protéines O-glycosylées :
liaison O-glycosidique entre l'hémiacétal d'un ose et l'hydroxyle d'un résidu Ser ou Thr



protéines N-glycosylées :
liaison N-glycosidique entre l'hémiacétal d'un ose et le -NH₂ de la fonction amide d'un résidu Asn.



Glycéro-glycolipides



Alditols

